

## PROJECT ADMINISTRATION DATA SHEET

☒ ORIGINAL ☐ REVISION NO. \_\_\_\_\_Project No. G-33-676 GTRI/CMX DATE 8 / 8 /84Project Director: Dr. Sheldon W. May School/CMX ChemistrySponsor: National Science FoundationType Agreement: Grant No. PCM-8402518Award Period: From 8/1/84 To 1/31/87 \* (Performance) 4/30/87 (Reports)Sponsor Amount: This Change Total to DateEstimated: \$                      \$ 93,000Funded: \$                      \$ 93,000Cost Sharing Amount: \$ 4,464 Cost Sharing No: G-33-355Title: "Enzymatic Epoxidation and Oxygen Activation"

## ADMINISTRATIVE DATA

OCA Contact

DAVID BRIDGES  
Lynn Boyd x4820

1) Sponsor Technical Contact:

2) Sponsor Admin/Contractual Matters:

Dr. Mary KirtleyMs. Ramona LaudaBiochemistry Program DirectorGrants OfficialNational Science FoundationNational Science FoundationWashington, DC 20550Washington, DC 20550(202) 357-7945(202) 357-9653

Defense Priority Rating: \_\_\_\_\_ Military Security Classification: \_\_\_\_\_

(or) Company/Industrial Proprietary: \_\_\_\_\_

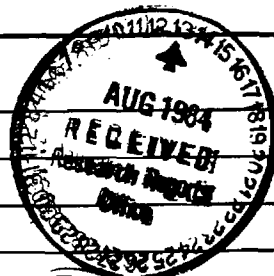
## RESTRICTIONS

See Attached NSF Supplemental Information Sheet for Additional Requirements.

Travel: Foreign travel must have prior approval – Contact OCA in each case. Domestic travel requires sponsor approval where total will exceed greater of \$500 or 125% of approved proposal budget category.

Equipment: Title vests with GIT

## COMMENTS:

\*includes usual 6-month unfunded flexibility period.

## COPIES TO:

Sponsor I.D. #02.107.000.84.037

Project Director  
Research Administrative Network  
Research Property Management  
AccountingProcurement/EES Supply Services  
Research Security Services  
✓ Reports Coordinator (OCA)  
Research Communications (2)GTRI  
Library  
Project File  
Other Newton

GEORGIA INSTITUTE OF TECHNOLOGY  
OFFICE OF CONTRACT ADMINISTRATION

NOTICE OF PROJECT CLOSEOUT

Closeout Notice Date 02/26/91

Project No. G-33-676 Center No. R5820-0A0

Project Director MAY S W School/Lab CHEMISTRY

Sponsor NATL SCIENCE FOUNDATION/GENERAL

Contract/Grant No. PCM-8402518 Contract Entity GTRC

Prime Contract No.

Title ENZYMATIC EPOXIDATION & OXYGEN ACTIVATION

Effective Completion Date 870131 (Performance) 870430 (Reports)

| Closeout Actions Required:                          | Y/N | Date Submitted |
|---|-----|----------------|
| Final Invoice or Copy of Final Invoice              | N   |                |
| Final Report of Inventions and/or Subcontracts      | Y   | 861125         |
| Government Property Inventory & Related Certificate | N   |                |
| Classified Material Certificate                     | N   |                |
| Release and Assignment                              | N   |                |
| Other   | N   |                |

Comments 98-A SATISFIES THE REQUIREMENT FOR THE PATENT REPORT, SUBMITTED WITH THE FINAL REPORT. NO INVOICE REQUIRED-NSF LOC.

Subproject Under Main Project No.

Continues Project No.

Distribution Required:

|                                       |   |
|---------------------------------------|---|
| Project Director                      | Y |
| Administrative Network Representative | Y |
| GTRI Accounting/Grants and Contracts  | Y |
| Procurement/Supply Services           | Y |
| Research Property Management          | Y |
| Research Security Services            | N |
| Reports Coordinator (OCA)             | Y |
| GTRC                                  | Y |
| Project File                          | Y |
| Other                                 | N |
|                                       | N |

NOTE: Final Patent Questionnaire sent to PDPI. 98A

Annual Technical Letter  
NSF Grant PCM-8402518

ENZYMATIC EPOXIDATION AND OXYGEN ACTIVATION

Sheldon W. May  
School of Chemistry  
Georgia Institute of Technology  
Atlanta, Ga

The objectives of this research program, focusing on the monooxygenase system from *P. oleovorans* which carries out epoxidation and hydroxylation of simple aliphatic hydrocarbon substrates (POEHS), are to extend and amplify the mechanistic information we have on the chemical pathway of the reaction, to extend our investigations to the design and evaluation of novel substrates, to focus on interactions at the active site critical to catalysis, and to carry out initial feasibility studies for applying physical techniques to this monooxygenase. The following paragraphs summarize our progress.

**Mechanistic and Stereochemical Studies:** Work with deuterio-olefins 5 has now been completed, and has confirmed the mechanistic view outlined in Scheme I of the proposal. Deuterium migration concomitant with aldehyde formation has now been fully confirmed by mass spectral analysis of products produced from 1,1-diD-1-octene. Furthermore, we successfully completed synthesis of cis-1-D-1-octene and confirmed the 70% inversion of configuration we reported with the trans-1-D-1-octene. Taken together, our finding of corresponding inversion of olefinic geometry together with the D migration accompanying aldehyde formation fully confirm our mechanistic hypothesis, and this work has now been published in a full paper in *J. Am. Chem. Soc.* Turning to fluorinated substrate analogs, synthesis of 1-F-1-octene, 1,1-diF-1-octene and 2-F-octene have been completed. Characterization of the enzymatic products formed from these substrates is currently being carried out by gc/ms, with CI ms proving essential since complex results are being obtained. We have preliminary evidence for formation of hydroxyoctanoate from both the 1,1-diF- and the 2-F substrates, but the sequence of chemical steps giving rise to this product is still unclear. Model studies on authentic fluorinated epoxides, possible immediate enzymatic products, are currently being carried out. Assay work with both 1-fluoro-octane and 1-H-perfluoro-octane 3 has been completed using the F electrode; both compounds are good substrates. Assay rates show linear dependence on monooxygenase present, absolute dependence on the presence of all components, and a sigmoidal dependence on FeRd, exactly as seen in the standard gc assays. A problem in this work has been the deactivating effect of F substitution which makes product identification studies much more difficult. With our discovery of striking activation of POEHS by imidazole, we now plan to reexamine the F compounds under conditions of much higher turnover. We have found that 1,7-octadiyne is a potent time-dependent inactivator of POEHS. Inactivation exhibits characteristics of mechanism-based irreversible inhibition, and further detailed studies on this and other mechanism-based inactivators are planned. An aspect of the work on these inactivators which we did not anticipate in the proposal is that with the various new catalytic competences we are now finding for POEHS (see below). Mechanism-based inactivators represent tools of choice for demonstrating that these quite distinct substrate classes all undergo reaction at a common active site.

Heteroatom-containing substrates: An extensive amount of work has been done in this area, far beyond what we were able to visualize when the proposal was written. The results have established several heretofore unknown activities for our prototypical NHI monooxygenase, POEHS. First, with thioether substrates (e.g. heptyl methyl sulfide) we have discovered that POEHS readily catalyzes oxygenative conversion to both sulfoxide and S-dealkylation products. Sulfide oxygenation exhibits the enzymological characteristics of the normal oxygenative pathway of POEHS, is kinetically facile, and is inhibited by our suicide substrate, 1,7-octadiyne. Studies with a series of thioether substrates revealed that partitioning between the sulfoxide and S-demethylation products is a function of substrate structure. Thus, octyl methyl sulfide is oxidized to the chiral sulfoxide, while p-methoxy phenethyl sulfide gives only the S-demethylation product. S-dealkylation is a process that has not previously been observed with monooxygenases, so we unequivocally demonstrated that this indeed proceeds via an oxygenative pathway by quantitative trapping of formaldehyde and the thiol; a simple displacement pathway would produce a C-1 product at the oxidation state of methanol. Several types of oxygen-containing substrate analogs have now been investigated. First, we find that terminal oxygenative O-demethylation (e.g. with 1-methoxyoctane) is very readily effected by POEHS, and this is the most kinetically facile activity we have found to date. Several lines of evidence establish that this reaction proceeds via the normal oxygenative POEHS pathway. Strikingly, we have also found that POEHS is indeed capable of non-terminal oxyfunctionalization, as illustrated by the production of either secondary alcohols or ketones via oxygenative O-demethylation of branched alkyl methyl or branched vinyl methyl ethers, respectively. Thus, for example, both 2- and 3-methoxy alkanes readily undergo O-demethylation to the 2- or 3- alcohols, and the vinyl ether, 2-OMe-1-octene, undergoes oxygenative ketonization. In all cases, as expected for an oxygenative process, formaldehyde is also produced stoichiometrically. In related work, we also discovered the facile oxygenation of terminal alcohols to aldehydes by POEHS, again carefully establishing all the enzymological characteristics of the reaction. In our view, these results are highly significant. We have now demonstrated the first example of chiral sulfoxidation by any enzyme system of this type. Coupled with our mechanistic and stereochemical results with olefinic substrates, analysis of the stereochemistry and mechanism of sulfur oxygenation will provide important insight into the chemistry of catalysis. Secondly, our discovery of the heretofore unrecognized competence of POEHS for oxygenation of non-terminal moieties significantly expands the menu of capabilities for this prototypic monooxygenase. This not only allows much more flexibility in the design of new substrates or inhibitors, but it also suggests a greatly expanded synthetic potential for this and other closely related bacterial hydrocarbon hydroxylating enzymes. Our working hypothesis for the mechanism of action of POEHS postulates that the substrate and binding sites of this enzyme are arranged in a manner necessitating oxygen attack at the terminal carbon. It now becomes clear that the reactivity of POM can be directed to positions other than the terminus of a straight chain, given an appropriate balance of binding and inherent reactivity of the functionality undergoing initial electron transfer to the active site metal.

Biochemistry Focusing on the Catalytic Site: Within the past few months we have worked out a new isolation procedure for POM which utilizes FPLC. This will facilitate eventual EXAFS studies where the most difficult biochemical impediment is the requirement for large amounts of enzyme at high concentration. As stated in our proposal, we will draw heavily on our experience with PAH in attempting to apply EXAFS to POEHS. An important finding which we have made recently is that POEHS catalysis is strikingly accelerated by imidazole, presumably via interaction with the active site iron. Stimulation is such that molecules

whose activities were heretofore unmeasurably slow can be clearly seen to undergo reaction. A similar observation has been recently reported by Groves and Watanabe with an iron porphyrin model epoxidation system. This obviously allows us to reexamine reactivity of molecules such as F-olefins which are inherently unreactive toward oxygenation. Characterization and scoping of this imidazole activation and its possible extension to other heterocycles represents an important goal for the future. Furthermore, since we can clearly pick out an imidazole ligand to Fe by EXAFS, as demonstrated in our published work with PCD, a goal in the future will be to see whether the imidazole binds directly to the Fe. If so, it will provide a handle for examining ligation and geometry changes through EXAFS.

School of Chemistry  
(404) 894-4002

**Georgia Institute of Technology**

Atlanta, Georgia 30332

A Unit of the University System of Georgia

November 25, 1986

Dr. Stella K. Engel  
Chemistry Program  
National Science Foundation  
Washington, D.C. 20550

84-02518

Dear Dr. Engel:

I have just been informed that the grants office at Georgia Tech has no record of the enclosed report, which I prepared last fall and sent thorough to them for forwarding to you. Therefore, I am sending the enclosed copy to you, and I am sure, in fact, you did not receive the original on account of some clerical error.

Best regards.

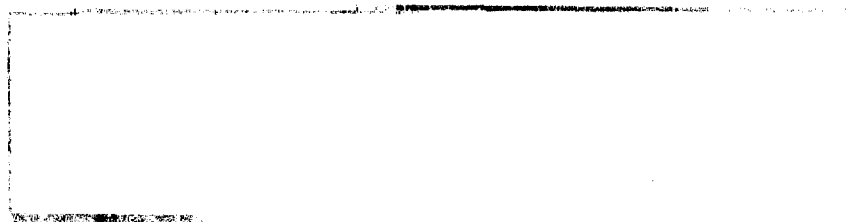
Very truly yours,

Sheldon W. May  
Professor of Chemistry

NATIONAL SCIENCE FOUNDATION  
 STREET, NW  
 WASHINGTON, DC 20550

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# NATIONAL SCIENCE FOUNDATION FINAL PROJECT REPORT

## PART I - PROJECT IDENTIFICATION INFORMATION

|                             |                           |     |  |
|-----------------------------|---------------------------|-----|--|
| Project Title               | Bionanotechnology Program |     |  |
| Principal Investigator      | Marlene Steinberg         |     |  |
| Project Number (NSF)        | From:                     | To: |  |
| Project Start and End Dates |                           |     |  |
| Project Location            |                           |     |  |
| Project Description         |                           |     |  |
| Project Objectives          |                           |     |  |
| Project Results             |                           |     |  |
| Project Conclusions         |                           |     |  |
| Project Significance        |                           |     |  |
| Project Impact              |                           |     |  |
| Project Future Plans        |                           |     |  |

This Packet Contains  
 NSF Form 98A  
 And 1 Return Envelope







# ANIMALS, EPIDEMIOLOGY AND OUTCOME ACTIVATION

Donald W. May  
School of Chemistry  
Georgia Institute of Technology  
Atlanta, Ga

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... on the chemical pathway of the reaction, to extend our ...  
... of novel substrates, to determine ...  
... critical to catalysis, and to carry out initial ...  
... studies for applying physical techniques to this ...  
... progress.

**Mechanistic and Stereochemical Studies:** Work with ...  
... has confirmed the mechanistic view outlined in ...  
... migration concomitant with aldehyde ...  
... mass spectral analysis of products produced from ...  
... Furthermore, we successfully completed synthesis of ...  
... and confirmed the  $20^\circ$  inversion of configuration ...  
... migration. Taken together, our finding of ...  
... together with the  $1^\circ$  migration accompanying ...  
... mechanistic hypothesis, and this work has ...  
... fluorinated substrates ...  
... 1,1-difluoro-1-octene and 2-F-octene have been ...  
... of the enzymatic products formed from these ...  
... with CI as proving ...  
... preliminary evidence for ...  
... and the 2-F substrate ...  
... is still unclear. ...  
... immediate enzymatic ...  
... with both 1-fluoro ...  
... using the F ...  
... dependence on monomer ...  
... of all components, and ...  
... gc assays. A ...  
... effect of F substitution which makes ...

... Mechanism-based ...  
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new class of ligands and geometry changes through EXAFS.